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### PHYTOLACCOSIDE B: TRITERPENE GLUCOSIDE FROM *PHYTOLACCA AMERICANA*

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**Key Word Index**—*Phytolacca americana*; Phytolaccaceae; poke-weed; triterpenoid glucoside; phytolaccoside B.

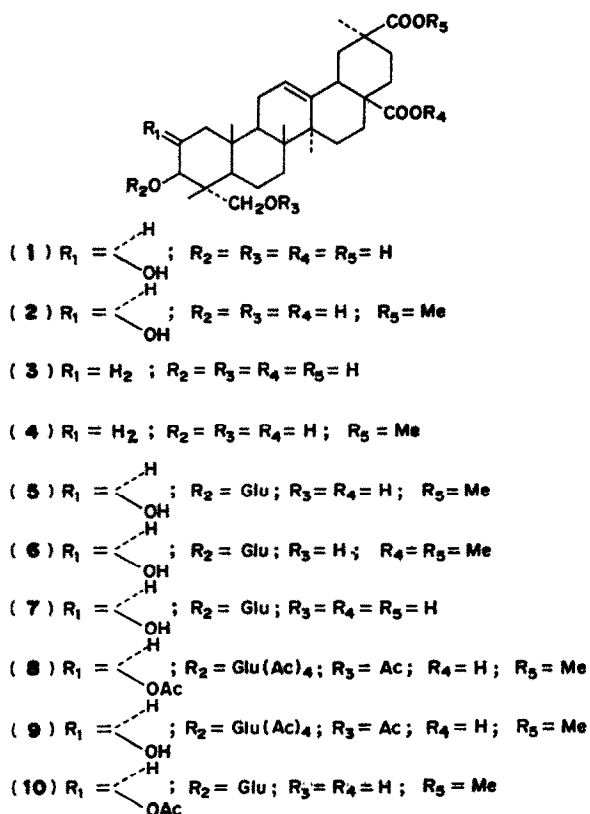
**Abstract**—The structure of phytolaccoside B, one of the major saponin components of the roots of *Phytolacca americana*, has been elucidated as 3 $\beta$ -D-glucopyranosyljaligonic acid 30-methyl ester.

The roots of *Phytolacca americana* are reputed in Korean medicine to alleviate rheumatism. They were found rich in saponins with strong anti-inflammatory activity [1]. According to earlier work [2,3] hydrolysis of the total saponin mixture yielded four sapogenins, jaligonic acid (1) [4] and its 30-methylester (2, phytolaccagenin) [5], and esculentic acid (3) [6] and its 30-methylester (4, phytolaccagenic acid) [7]. This paper describes the structural elucidation of phytolaccoside B, one of the major saponins isolated from the roots of this plant.

Phytolaccoside B (5),  $C_{37}H_{58}O_{12}$ , mp 215–218°,  $[\alpha]_D^{25} + 75.8^\circ$ , on acid hydrolysis, gave jaligonic acid 30-methylester (2), mp 317–319°, as the genin, which was identified by direct comparison with an authentic sample (mp, TLC, IR and MS). Glucose was identified in the hydrolysate from the saponin (5) by TLC. The yield of the genin in a quantitative hydrolysis experiment showed the presence of one sugar unit in the molecule (genin found 73%; calc. 76.66%).

Methylation of 5 with  $CH_2N_2$  gave methylated product 6, mp 182–184°,  $[\alpha]_D^{25} + 71.6^\circ$ . Saponification of 5 with alkali afforded a new glucoside 7, mp 234–239°,  $[\alpha]_D^{25} + 77^\circ$ , which gave 6 by methylation with  $CH_2N_2$ . These results show that the sugar must be present in glucosidic linkage with one of the hydroxyl groups in ring A of the genin (not in ester linkage with the 28-carboxyl group).

In a quantitative periodate oxidation experiment 2 moles of the reagent were consumed. This indicates that the sugar is attached to the C-2 or C-3 position of the genin.



(11)  $R_1 = O$  ;  $R_2 = R_3 = R_4 = H$  ;  $R_5 = Me$

(12)  $R_1 = \begin{array}{c} \text{H} \\ \diagup \\ \text{C} \\ \diagdown \\ \text{OH} \end{array}$  ;  $R_2 = R_3 = Ac$  ;  $R_4 = H$  ;  $R_5 = Me$

(13)  $R_1 = O$  ;  $R_2 = Glu(Ac)_4$  ;  $R_3 = Ac$  ;  $R_4 = H$  ;  $R_5 = Me$

Acetylation of **5** with acetic anhydride-pyridine on a steam bath afforded two compounds, a hexaacetate (**8**), mp 163–166°,  $[\alpha]_D^{24} + 58.8^\circ$ , and a pentaacetate (**9**), mp 171–174°,  $[\alpha]_D^{24} + 62.3^\circ$ , in a 7:3 ratio. When acetylated at 4°, the major product was **9**. Saponification of **9** with  $NH_3$  gave the original saponin (**5**), however a monoacetate (**10**), mp 206–210°,  $[\alpha]_D^{26} + 69.2^\circ$ , was formed from **8** under the same conditions. This result suggests that the more sterically hindered axial secondary alcohol group (i.e. 2 $\beta$ -OH) of the genin in the saponin (**5**) is free and therefore the sugar is linked to the 3 $\beta$ -OH group.

This suggestion was supported by Sarett oxidation of **9** followed by hydrolysis to yield 2-oxo-esculentic acid 30-methylester (**11**), mp 222–224°,  $\lambda_{max}$  204 and 282 nm (log  $\epsilon$  3.67 and 2.63), which was identified by direct comparison with a sample prepared by oxidation of 3,23-diacetyl jaligonic acid 30-methylester (**12**), followed by hydrolysis.

The NMR spectra of **8**, **9** and **13**, in which the one proton doublets were observed at  $\delta$  4.52 ( $J$  7 Hz), 4.50 ( $J$  7.5 Hz) and 4.49 ( $J$  6 Hz), respectively, indicated the  $\beta$ -configuration of the glucosidic linkage in **5**. This was also proved by Klyne's rule [8] (Molecular rotation difference between **2** and **5** =  $-84^\circ$ ,  $\beta$ -methyl-D-glucopyranoside =  $-62^\circ$ ). Thus the structure of phytolaccoside B was established as 3 $\beta$ -D-glucopyranosyljaligonic acid 30-methylester.

#### EXPERIMENTAL

Mp's are uncorrected. Optical rotations were measured in MeOH. IR spectra were recorded in KBr and UV spectra in EtOH. NMR spectra were taken in  $CDCl_3$  using TMS as internal reference at 100 MHz.

**Isolation of phytolaccoside B (5).** The saponin mixture, obtained from a MeOH extract of *P. americana* roots as described previously [2], showed 10 spots on TLC (MeOH- $CHCl_3$ - $NH_4OH$ - $H_2O$  30:60:3:4). These were designated alphabetically in order of decreasing  $R_f$  values. The mixture was applied to a Si gel column and eluted with MeOH- $CHCl_3$  (3:97) to give a fraction which contained only two compounds, phytolaccoside A and B. Phytolaccoside B (**5**) was obtained by PLC, mp 215–218°;  $[\alpha]_D^{25} + 75.8^\circ$  (MeOH;  $c$  0.546); IR  $\nu_{max}^{KBr}$   $cm^{-1}$ : 3340 (OH), 2900 (Me,  $CH_2$ ), 1725 and 1150 (methylester), 1700 (acid), 1000–1100 (glucoside); (Found: C, 63.89; H, 8.68.  $C_{37}H_{58}O_{12}$  requires: C, 63.96; H, 8.41%). Lieberman-Burchard and Molish tests were positive.

**Hydrolysis.** The saponin (150 mg) was heated with 5% HCl-MeOH under reflux for 2 hr. The product was diluted with  $H_2O$  and extracted with  $Et_2O$ . The  $Et_2O$  extract was crystallized from MeOH to give needles of **2** (110 mg), mp 317–319°, which was identified by direct comparison with an authentic sample. The aq. part was examined by TLC (MeOH- $CHCl_3$ - $Me_2CO$ - $NH_4OH$  5:2:3:2) and it showed the presence of D-glucose only.

**Methylation.** The saponin (50 mg) in MeOH (20 ml) was treated with  $CH_2N_2$  in the usual manner. The crude product was purified by PLC ( $CHCl_3$ -MeOH- $NH_4OH$ - $H_2O$  20:4:1:3) and crystallized from MeOH to give needles of **6**, mp 182–184°;  $[\alpha]_D^{25} + 71.6^\circ$  (MeOH;  $c$  0.725); IR  $\nu_{max}^{KBr}$   $cm^{-1}$ : 1725, 1150 (methylester); (Found: C, 63.95; H, 8.78.  $C_{38}H_{60}O_{12}$  requires: C, 64.39; H, 8.53%).

**Saponification.** The saponin (100 mg) was heated under reflux in 20 ml of 3% NaOH-MeOH for 3 hr. The reaction mixture was acidified and extracted with BuOH. The BuOH extract was crystallized from MeOH to yield needles of **7**, mp 234–239°;  $[\alpha]_D^{25} + 77^\circ$  (MeOH;  $c$  0.5); (Found: C, 63.44; H, 8.15.  $C_{36}H_{56}O_{12}$  requires: C, 63.51; H, 8.29%). Methylation of **7** with  $CH_2N_2$  gave **6**.

**HIO<sub>4</sub> oxidation.** A MeOH soln of the saponin (34.8 mg) was added to 0.01 M HIO<sub>4</sub> (20 ml), the volume made up to 35 ml with MeOH and the soln kept in the dark at 4°. Aliquots (7 ml each) were removed after 2, 4, 5, 28 and 35 hr. Estimation of the HIO<sub>4</sub> with 0.01 M  $Na_3AsO_3$  in the usual way showed the consumption of 1.54, 1.85, 1.95, 1.98 and 1.98 mol of oxidant, respectively.

**Acetylation.** The saponin (100 mg) was heated with  $Ac_2O$  (3 ml) and  $C_5H_5N$  (2 ml) for 3 hr. Pouring onto ice gave solids which were chromatographed on Si gel and elution with MeOH- $CHCl_3$  (1:99) yielded **8** (65 mg) in the earlier fractions, mp 163–166°;  $[\alpha]_D^{24} + 58.8^\circ$  (MeOH;  $c$  1.0); NMR:  $\delta$  0.74–1.16 (5  $\times$  Me), 2.05–2.13 (6  $\times$  MeCO), 3.72 ( $CH_3O$ ), 4.52 (1H,  $d$ ,  $J$  7.0 Hz, anomeric proton); (Found: C, 62.21; H, 7.47.  $C_{49}H_{70}O_{18}$  requires: C, 62.14; H, 7.45%). From the later fractions, **9** was obtained, mp 171–174°;  $[\alpha]_D^{24} + 62.3^\circ$  (MeOH;  $c$  1.0); NMR:  $\delta$  0.71–1.24 (5  $\times$  Me), 2.04–2.09 (5  $\times$  MeCO), 3.69 (MeO), 4.50 (1H,  $d$ ,  $J$  7.5 Hz, anomeric proton); (Found: 62.31; H, 7.58.  $C_{47}H_{68}O_{17}$  requires: C, 62.37; H, 7.57%). On 10 hr treatment **9** was completely acetylated to give **8**. Partial acetylation of the saponin with  $Ac_2O$ - $C_5H_5N$  at 4° overnight gave predominantly **9**.

**Mild hydrolysis of saponin acetates.** A sample (50 mg) of **8** in MeOH (7 ml) and  $NH_4OH$  (7 ml) was kept at 50° for 12 hr. The reaction mixture was concentrated under red pres and chromatographed on Si gel. Elution with MeOH- $CHCl_3$  (3:97) gave **10**, mp 206–210°;  $[\alpha]_D^{26} + 69.2^\circ$  (MeOH;  $c$  0.5); (Found: C, 63.30; H, 8.22.  $C_{39}H_{60}O_{13}$  requires: C, 63.57; H, 8.21%). On deacetylation under the same conditions, **9** yielded **5**.

**Sarett oxidation of the pentaacetate (9).** A sample (100 mg) of **9** in  $C_5H_5N$  (3 ml) was added to  $CrO_3$  (100 mg) in  $C_5H_5N$  (2 ml) and left overnight at room temp. Since the oxidation product could not be separated from the unreacted compound, the mixture was acetylated to convert **9** into **8** and oxidised product (**13**) was separated by means of PLC ( $CHCl_3$ -MeOH- $NH_4OH$ - $H_2O$  20:4:1:3), mp 155–157°;  $[\alpha]_D^{25} + 39.5^\circ$  (MeOH;  $c$  0.66); NMR:  $\delta$  0.71–1.18 (5  $\times$  Me), 2.06–2.15 (5  $\times$   $CH_3CO$ ), 3.69 ( $CH_3O$ ), 4.19 (1H,  $s$ , C-3), 4.49 (1H,  $d$ ,  $J$  6 Hz, anomeric proton); (Found: C, 62.49; H, 7.33.  $C_{47}H_{66}O_{17}$  requires: C, 62.51; H, 7.37%). On acid hydrolysis **13** yielded the genin, mp 222–224°, which was identical with 2-oxo-esculentic acid 30-methylester (**11**) prepared as described below.

**Synthesis of 2-oxo-esculentic acid 30-methylester (11).** Partial acetylation of **2** with  $Ac_2O$ - $C_5H_5N$  at 4° overnight gave 3,23-diacetyl jaligonic acid 30-methylester (**12**), mp 170–174°;  $[\alpha]_D^{24} + 100^\circ$  (MeOH,  $c$  0.496); NMR:  $\delta$  0.74–1.33 (5  $\times$  Me), 2.10 (3H,  $s$ , MeCO), 2.18 (3H,  $s$ , MeCO), 3.72 (3H,  $s$ , MeO), 4.24 (1H,  $m$ , C-2), 4.49 (1H,  $d$ ,  $J$  3 Hz, C-3) and 5.37 (1H,  $m$ , C-12); (Found: C, 68.02; H, 8.45.  $C_{35}H_{52}O_9$  requires: C, 68.16; 8.50%). A sample (70 mg) of **12** was oxidized with  $CrO_3$ - $C_5H_5N$  in the usual way. The reaction product was deacetylated by refluxing in 3% HCl-MeOH, and chromatographed over Si gel. Elution with MeOH- $CHCl_3$  (2:98) gave **11** which was crystallized from  $Et_2O$  as flakes, mp 222–224°; UV  $\lambda_{max}$  nm (log  $\epsilon$ ) 204 (3.67), 282 (2.63); (Found: C, 70.27; H, 8.74.  $C_{31}H_{46}O_7$  requires: C, 70.16; H, 8.74%).

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# NIC-1-LACTONE, A MINOR STEROIDAL CONSTITUENT OF *NICANDRA PHYSALOIDES* (SOLANACEAE)

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**Key Word Index**—*Nicandra physaloides*; Solanaceae; aromatic rang D; steroid; withanolides; Nic-1-lactone.

We wish to report on the isolation and characterization of Nic-1-lactone  $C_{28}H_{32}O_6$ , **1a**, mp 230–232°, a minor steroidal constituent of *Nicandra physaloides* (Solanaceae), the only plant known [1,2] to contain steroids with an aromatic D ring. The compound was found to be identical with 6 $\alpha$ ,7 $\alpha$ :24,25-diepoxy-5-hydroxy-17(13→18)abeo-5 $\alpha$ -ergosta-2,13,15,17-tetraen-1-one-26,22-lactone, obtained [3] by oxidation with chromium trioxide of the lactolic C-26 hydroxyl group in Nic-1 (**1b**), the major steroidal component of *N. physaloides*.

The Nic constituents may be divided into two groups: (a) those with the usual steroid skeleton as encountered in Nic-3 (**2a**), Nic-7 (**2b**) [1] and withanicandrin (**2c**) [4]; (b) those with a rearranged skeleton, as found in Nic-1 (**1b**) [1,2], obtained by expansion of ring D through inclusion of the C-13 methyl and subsequent aromatization. Most Nic constituents characterized so far possess an ergostane type side chain which has undergone oxida-

tive processes leading to the closure of a six membered epoxy-lactol. In several compounds this side chain has gradually lost most of its carbon atoms, until a two carbon fragment remains as present in Nic-10 (**1c**) [1]. The usual unsaturated six membered ring lactone, characteristic for the majority of the withanolides [4], is found in only one compound withanicandrin (**2c**), occurring in this plant [4].

In our opinion, the epoxy-lactol system present in Nic-1-lactone (**1a**) is formed in Nature by oxidation of the lactolic hydroxyl group present in Nic-1 (**1b**) and not by epoxidation of the  $\Delta^{24}$  double bond of an unsaturated lactone such as that present in the side chain of withanicandrin (**2c**). This assumption is based on the fact that all the solanaceous plants which produce withanolides, Nic-derivatives and physalins, possess enzymes which can catalyse the epoxidation of isolated double bonds, or double bonds in allylic systems, but which cannot epoxidize double bonds in  $\alpha\beta$ -unsaturated carbonyl systems.

## EXPERIMENTAL

*N. physaloides* was raised in the nursery of the Volcani Center, Bet Dagon, Israel (introduction No. 20-707), from seeds received from Pretoria, South Africa (No. M-71-384). It is interesting to note that populations of plants raised from seeds received from India (Pondicherry) did not contain compound **1a**.

The isolation was done according to the described procedure [4]. The first fractions containing Nic-1, obtained by chromatography on a column of Si gel H, were contaminated by a product possessing a slightly higher  $R_f$  value. Separation was achieved by PLC on Si gel PF<sub>254</sub>, 1 mm thickness, 40 cm length, developed with  $C_6H_6$ -EtOAc (1:4). The product possessing the higher  $R_f$  value (accounting for only 5% of the total amount of Nic-1) crystallised from  $Me_2CO$ -hexane, mp 230–232°, undepressed on admixture with 6 $\alpha$ ,7 $\alpha$ :24,25-diepoxy-5-hydroxy-17(13→18)abeo-5 $\alpha$ -ergosta-2,13,15,17-tetraen-1-one-26,22-lactone [3]. All physical constants and spectral data were superimposable.

